



CKJ REVIEW

Sweet dreams: therapeutic insights, targeting imaging and physiologic evidence linking sleep, melatonin and diabetic nephropathy

Baris Afsar¹, Rengin Elsurer Afsar¹, Alan A. Sag², Asiye Kanbay³, Hakan Korkmaz⁴, José Cipolla-Neto⁵, Adrian Covic⁶, Alberto Ortiz⁷ and Mehmet Kanbay ⁸

¹Division of Nephrology, Department of Medicine, Suleyman Demirel University School of Medicine, Isparta, Turkey, ²Division of Vascular and Interventional Radiology, Department of Radiology, Duke University Medical Center, Durham, NC, USA, ³Department of Pulmonary Medicine, Istanbul Medeniyet University School of Medicine, Istanbul, Turkey, ⁴Division of Endocrinology, Department of Medicine, Suleyman Demirel University School of Medicine, Isparta, Turkey, ⁵Institute of Biomedical Sciences, University of Sao Paulo, Sao Paulo, Brazil, ⁶Dialysis Unit, School of Medicine, IIS-Fundacion Jimenez Diaz, Universidad Autónoma de Madrid, Madrid, Spain, ⁷Nephrology Clinic, Dialysis and Renal Transplant Center, 'C.I. PARHON' University Hospital and 'Grigore T. Popa' University of Medicine, Iasi, Romania and ⁸Division of Nephrology, Department of Medicine, Koç University School of Medicine, Istanbul, Turkey

Correspondence to: Mehmet Kanbay; E-mail: drkanbay@yahoo.com; mkanbay@ku.edu.tr

ABSTRACT

Melatonin is the main biochronologic molecular mediator of circadian rhythm and sleep. It is also a powerful antioxidant and has roles in other physiologic pathways. Melatonin deficiency is associated with metabolic derangements including glucose and cholesterol dysregulation, hypertension, disordered sleep and even cancer, likely due to altered immunity. Diabetic nephropathy (DN) is a key microvascular complication of both type 1 and 2 diabetes. DN is the end result of a complex combination of metabolic, haemodynamic, oxidative and inflammatory factors. Interestingly, these same factors have been linked to melatonin deficiency. This report will collate in a clinician-oriented fashion the mechanistic link between melatonin deficiency and factors contributing to DN.

Keywords: diabetes, diabetic nephropathy, inflammation, kidney disease, melatonin, oxidative stress

INTRODUCTION

Melatonin is an indolamine that is present in almost every organism from bacteria to humans [1]. In mammals, the site of

hormonal melatonin production is the pineal gland, but melatonin is also produced in peripheral tissues for local autocrine and paracrine actions. Pineal melatonin production is restricted to the night and its production duration follows the duration of

Received: 19.8.2019; Editorial decision: 16.12.2019

© The Author(s) 2020. Published by Oxford University Press on behalf of ERA-EDTA.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

the night. Melatonin mainly regulates biological rhythms and has a role in coordinating behavioural and physiological adaptations to the night/day cycle and seasons [2].

Diabetic nephropathy (DN) is a microvascular complication of diabetes and is the leading cause of renal failure. Blood pressure regulation, glycaemic control and management of hyperlipidaemia are still the mainstays of therapy. These have not resulted in a cure [3, 4]. Melatonin-based therapy may be another pathway for therapeutic synergy.

DN is driven by the metabolic derangement causing haemodynamic changes, oxidative stress and inflammation. In early stages, DN is characterized by glomerular hyperfiltration and podocyte loss [5, 6]. While melatonin deficiency causes metabolic derangements, haemodynamic changes, oxidative stress and inflammation, the potential nephroprotective effects of melatonin are understudied. In this review we summarized the current literature about the effect of melatonin on the development of DN and the underlying pathophysiology.

Melatonin synthesis is tied to light and the day/night cycle

Melatonin (N-acetyl-5-methoxy tryptamine) is a tryptophan-derived small molecule showing pleiotropic actions, including antioxidant properties [7]. In mammals, melatonin is centrally produced by the pineal gland, acting as a hormone and, in addition, melatonin is produced in several central and peripheral tissues (e.g. retina, astrocytes, gastrointestinal tract, bone marrow, lymphocytes and skin) where it acts as a paracrine/autocrine factor [8, 9].

Melatonin secretion is tightly regulated (Figure 1). Pineal gland melatonin strictly with nocturnal production depends on two factors: first, circadian timing by the suprachiasmatic hypothalamic nucleus, and second, the nocturnal production is restricted to the night due to the so-called photoinhibition of its production by light acting through the retinal melanopsinergic system originating in the intrinsic photosensitive ganglion cells [10]. However, in spite of being produced only during the night and in the dark, melatonin effects might be expressed not only during the night (immediate effects) but also during the day when melatonin is no longer circulating (prospective effects) [2]. Superior cervical ganglia provide sympathetic innervation to the pineal gland, releasing norepinephrine that stimulates the rate-limiting steps that convert tryptophan to melatonin in the pineal gland [2, 11]. Melatonin is not stored but is immediately released into the bloodstream and cerebrospinal fluid, bathing the brain and organs simultaneously. It has a short (40-min) half-life and is metabolized in the liver and kidneys and excreted renally as 6-sulfatoxymelatonin [12].

Melatonin activates two kinds of G-protein-linked membrane receptors, MT₁ (high affinity) and MT₂ (low affinity), which are encoded by the *MTNR1A* and *MTNR1B* genes, respectively. These receptors are expressed in multiple tissues such as heart and arteries, adrenal gland, kidney, lung, liver, gallbladder, small intestine, adipocytes, ovaries, uterus, breast, prostate, skin and central nervous system. They are also expressed by T and B lymphocytes [13]. However, receptor-expressing cells and tissues are not the only targets of melatonin physiologic actions since melatonin expresses non-receptor-dependent mechanisms of action such as, e.g., the direct nitrogen and oxygen radical species chelating antioxidant effects. As an antioxidant, melatonin protects DNA from oxidative damage [14–17], especially from mitochondrion-derived free radicals [18]. Melatonin also regulates ubiquitin-linked proteasomes to inhibit Ca²⁺/

calmodulin-dependent protein kinase II activity and decreases protein catabolism [19]. It additionally activates extracellular signal-regulated kinase and G-protein q subunit signalling [19].

Central and peripheral effects of melatonin

Melatonin regulates the circadian sleep–wake and body temperature cycles [20, 21]. This chronobiologic effect involves the hypothalamic suprachiasmatic nucleus as imaged by magnetic resonance imaging [22, 23].

The metabolic role of melatonin has been studied in rats, where pinealectomy leads to increased body weight owing to increased food intake and reduced energy expenditure [24]. Replacing melatonin in these rats reduced body weight and food intake and increased brown fat activation [25, 26]. Interestingly, post-menopausal women taking daily melatonin supplementation in a randomized placebo-controlled trial reduced fat mass and increased lean mass [27]. In addition to these indirect antidiabetic effects, melatonin directly increases pancreatic beta cell survival and function [28–30] by increasing insulin secretion through glucagon-like peptide-1 sensitization [31]. In a population-based study, lower melatonin levels were independently associated with the risk of developing type 2 diabetes, possibly because melatonin regulates glucose tolerance [2, 32–34], and of insulin release in a complex feedback loop [35, 36].

Melatonin also regulates haemodynamic equilibrium. Pinealectomized rats became hypertensive, and this was resolved with melatonin supplementation [37]. Separately, 24-h light exposure (and the resultant melatonin suppression) causes hypertension via sympathetic drive and renin–angiotensin system activation (vasoconstriction and volume retention) [37, 38]. These mechanisms are activated by cardiovascular system melatonin receptors [39]. Also, melatonin acts on mitochondria regulation to maintain a healthy cardiovascular system [40]. In addition, direct brain actions of melatonin also reduce sympathetic tone and downregulate adrenal gland activity via the hypothalamus [41, 42]. Melatonin also modulates the baroreflex set point [43] and regulates heart rate via the medulla [44] and vasoconstriction and vasodilation via direct activation of vessel melatonin receptors [39, 45, 46]. In this regard, melatonin deficiency leads to blood pressure non-dipping or reverse dipping at night [47, 48]. In summary, because melatonin has cardiovascular and metabolic effects, derangements can result in diabetes and obesity (Figure 2).

DN occurs inconsistently and shortens lifespan

Technically, DN is defined as decreased glomerular filtration rate (GFR) and/or elevated urinary albumin excretion (30–300 mg/day microalbuminuria, >300 mg/day macroalbuminuria). Not all diabetics develop DN, but the reasons are unclear. Type 2 diabetics are more likely than type 1 diabetics to develop DN, although there are confounders such as older age and more frequent cardiovascular disease and atherosclerosis [49]. In any case, DN increases the risk of death in both type 1 and type 2 diabetics [50, 51] and ultimately progresses to end-stage kidney disease requiring renal replacement therapy by dialysis or transplantation [52]. However, albuminuria is inconsistently associated with a DN progression and some patients progress without albuminuria [53]. DN biopsies show a variety of pathologic findings involving almost every portion of the nephron, notably basement membrane thickening, podocyte loss and interstitial fibrosis [49]. A key pathogenic pathway is hyperglycaemia increasing

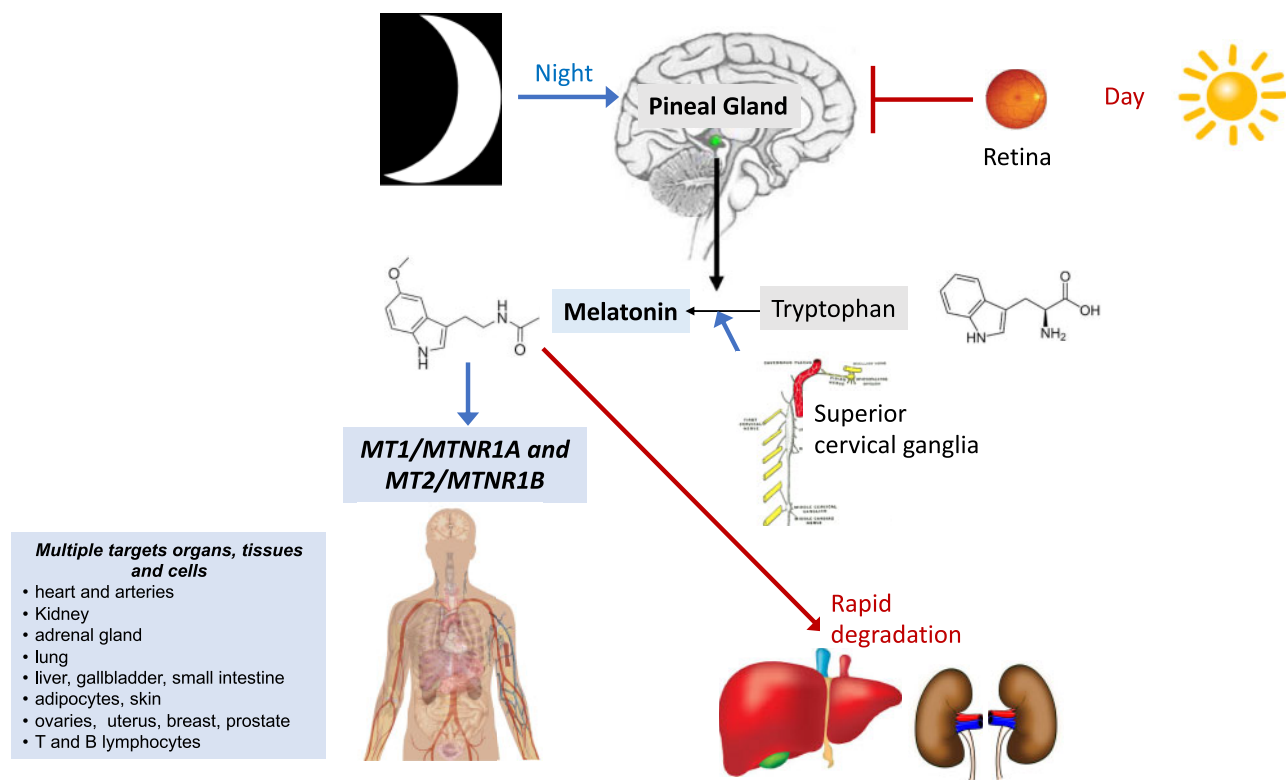


FIGURE 1: Melatonin secretion is regulated by diurnal rhythms of the body. Its secretion is increased during sleep and decreased during the daytime. Its systemic effects are regulated by activating MT1 (high affinity)/MT2 (low affinity) receptors. It is degraded in the liver and kidney.

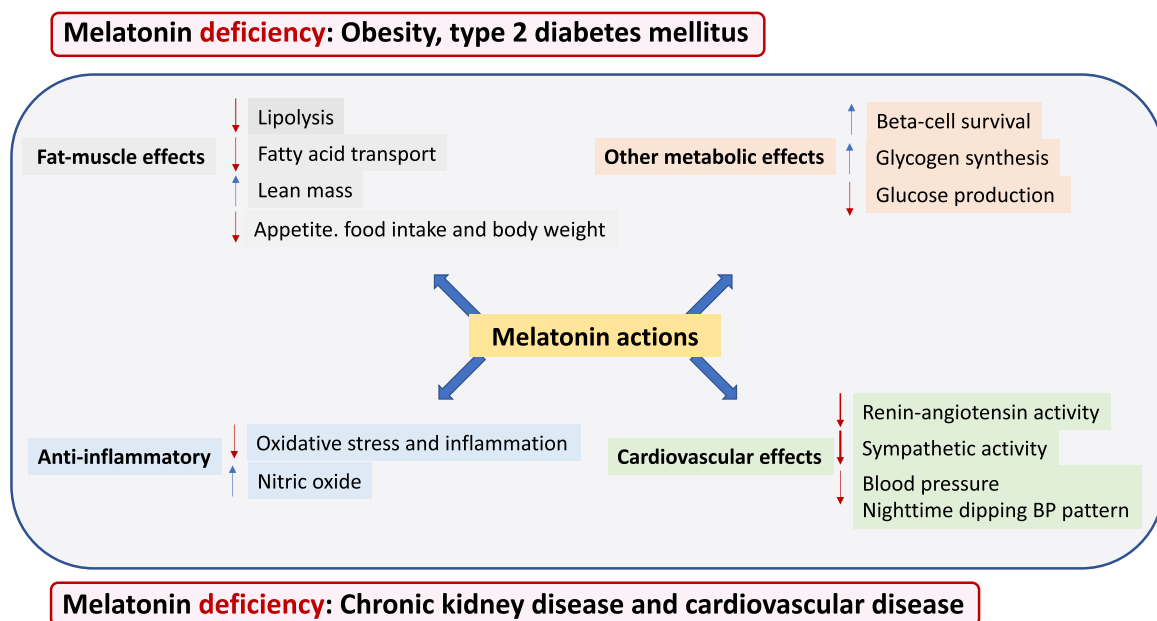


FIGURE 2: Melatonin deficiency may increase the risk for development of diabetes, obesity, cardiovascular and kidney disease. Melatonin has multiple health benefits on multiple organs.

mitochondrial substrate oxidation [49] to activate the reduced form of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, thus uncoupling nitric oxide synthase [54], resulting in the generation of reactive oxygen species (ROS). Excess ROS causes cell dysfunction, apoptosis and inflammation, and decreasing ROS exposure is beneficial [49, 55].

Other factors contributing to chronic kidney disease (CKD) progression include hypertension and impaired autoregulation, leading to hypoperfusion and inappropriate renin-angiotensin system activation [56] in both type 1 and 2 diabetes [57, 58]. Loss of renal autoregulation allows systemic hypertension to directly hit the glomerulus [59, 60]. Glucose-mediated endothelial

dysfunction promotes microvascular rarefaction and renal hypoxia [49, 61]. Not surprisingly, lowering blood pressure is protective in hypertensive DN [62–64]. Insulin resistance is linked to CKD [65, 66]. Thus insulin receptor deletion in podocytes leads to glomerular damage similar to that observed in DN [67]. Compensatory insulin hypersecretion promotes kidney fibrogenesis through actions in insulin-responsive cells, further contributing to progressive renal disease [68]. Obesity independently promotes inflammation and growth factor activity, thus promoting CKD progression [49]. In this regard, inappropriate recruitment of activated T cells and macrophages favours glomerular and tubulointerstitium inflammation and DN progression [69, 70]. Therapeutic approaches targeting inflammatory mediators decrease albuminuria and GFR loss in animals and humans with DN [71, 72].

Melatonin measurement in diabetes

Melatonin levels are known to vary in a diurnal pattern, with secretion in humans occurring mostly at night. Interestingly, the complications of diabetes impair this secretion. Retinal perception of light may disturb melatonin dynamics in patients with diabetic retinopathy. Autonomic neuropathy may impair innervation of pinealocytes, which leads to altered melatonin haemodynamics in diabetes. These diabetic consequences are less discussed than the common cardiovascular and lower extremity peripheral vascular consequences [73].

Hikichi *et al.* [74] compared both the night- and daytime melatonin secretion in non-diabetic and diabetic subjects. They found that diabetics had lower melatonin at night but daytime levels were not affected by diabetes. In another study, Tutuncu *et al.* [73] designed a study to determine melatonin dynamics in type 2 diabetic patients and its relationship with the autonomic nervous system. They measured melatonin levels between 2 and 4 a.m. and 4 and 6 p.m. and compared these in 36 diabetics versus 13 non-diabetics. Again, like with Hikichi *et al.*, diabetics had lower nighttime melatonin levels and less of a melatonin surge into nighttime, both statistically significant findings. Patients carrying a diagnosis of autonomic neuropathy showed lower night- and daytime melatonin levels compared with non-diabetics (both statistically significant). Retinopathy did not affect the findings but the authors suggested that the participants' degree of retinopathy was not severe enough to generate a signal [73]. Prior to these studies, O'Brien *et al.* [75] had already shown that a physiological increase in nocturnal plasma melatonin concentration is not observed in diabetic patients with neuropathy compared with age-matched non-diabetic controls. The compilation of studies supports the hypothesis that melatonin dysregulation is a novel diabetic complication. Future studies may focus on melatonin dynamics graded by the severity of diabetic neuropathy.

Melatonin and DN

Sleep patterns are linked to diabetes via insulin resistance and metabolic syndrome [76, 77] and the disturbed sleep–diabetes link [78] is likely driven by melatonin deficiency [79]. In fact, type 2 diabetics with decreased sleep had higher 24-h urinary albumin and protein excretion as markers of more severe DN [80]. Moreover, diabetes-derived hyperglycaemia induces a reduction in melatonin production, aggravating sleep and metabolic medical conditions [81].

Peschke *et al.* [82] showed that serial nocturnal plasma melatonin levels were significantly lower in six diabetic patients

compared with five non-diabetic controls. Although this study involves only a small number of patients, the performance of serial measurements improves the validity of the study [82]. Melatonin levels also vary with microvascular diabetic complications. Nocturnal plasma melatonin levels were studied in 56 patients by Hikichi *et al.* [74]. Interestingly, they found that the patients with diabetic proliferative retinopathy had lower melatonin levels than healthy patients. However, non-retinopathy diabetics did not demonstrate this finding. Kor *et al.* [83] compared the melatonin levels in 40 type 1 diabetic children and 30 non-diabetic controls. The mean melatonin level in the diabetic group was 6.75 ± 3.52 pg/mL and the mean melatonin level in the control group was 11.51 ± 4.74 pg/mL ($P < 0.01$). In their relatively small cross-sectional study, Robeva *et al.* [84] showed that nocturnal insulin and plasma melatonin levels correlated positively in metabolic syndrome patients but not healthy control patients. Melatonin deficiency may predispose to DN via vasoactive, metabolic, inflammatory, apoptotic and fibrogenic pathways (Figure 3).

Activation of Rho-associated kinases promotes endothelial-mesenchymal transformation [85–87] and DN progression, which is prevented by inhibiting this pathway [88]. In cultured cells, microRNA 497 attenuated Rho-associated kinase signalling [89]. Mesenchymal stem cell therapy improved renal function in rat DN and melatonin improved renal recovery by increasing antioxidant defences and decreasing immune activation [90].

Melatonin also modulates renin-angiotensin system activation, in general, and particularly in DN [91–93]. Thus the renin-angiotensin system was upregulated in CKD patients with impaired melatonin secretion at night [94]. In subtotaly nephrectomized rats, treatment with melatonin for 4 weeks improved remnant kidney function and decreased intrarenal renin-angiotensin activation and interstitial fibrosis [95]. In cultured cells, melatonin reduced the expression of apoptotic proteins in response to a diabetic milieu, resulting in increased podocyte numbers. Melatonin prevented angiotensin-2-driven pro-apoptotic protein transcription and protected mitochondrial membranes in a dose-dependent manner [96]. In rats with streptozotocin-induced DN, the combination of melatonin and taurine decreased glomerular inflammation and proteinuria, independent of serum glucose levels [97]. In the same model, melatonin also increased nitric oxide availability and nephroprotective protein levels, including those of antioxidant proteins such as superoxide dismutase [98], and also decreased kidney cell apoptosis [99], improving histological kidney damage [100]. Nephroprotection by melatonin is not limited to DN, but extends to potential clinical complications of diabetic patients. Thus melatonin reduced the inflammation marker interleukin-33 (IL-33) in streptozotocin-induced DN rats with contrast-induced nephropathy [101] and protected against adriamycin-induced podocytopathy [102]. It additionally inhibited and normalized NADPH oxidase activity, a key driver of oxidative stress that is upregulated in obese Zucker diabetic rats [103–105].

Macrophages are the predominant kidney infiltrating cells in DN [106, 107] and macrophage infiltration in biopsy specimens predicts GFR loss in DN [108]. Therapeutic manoeuvres that decrease macrophage infiltration also decrease albuminuria and slow DN regression [109, 110]. The nuclear factor κ B (NF- κ B) transcription factor is a master regulator of inflammation, contributing to DN progression by promoting macrophage recruitment and activation [111]. Macrophages secrete transforming growth factor β 1, a pro-fibrotic factor that plays a key role in DN-

Melatonin and Diabetic Nephropathy

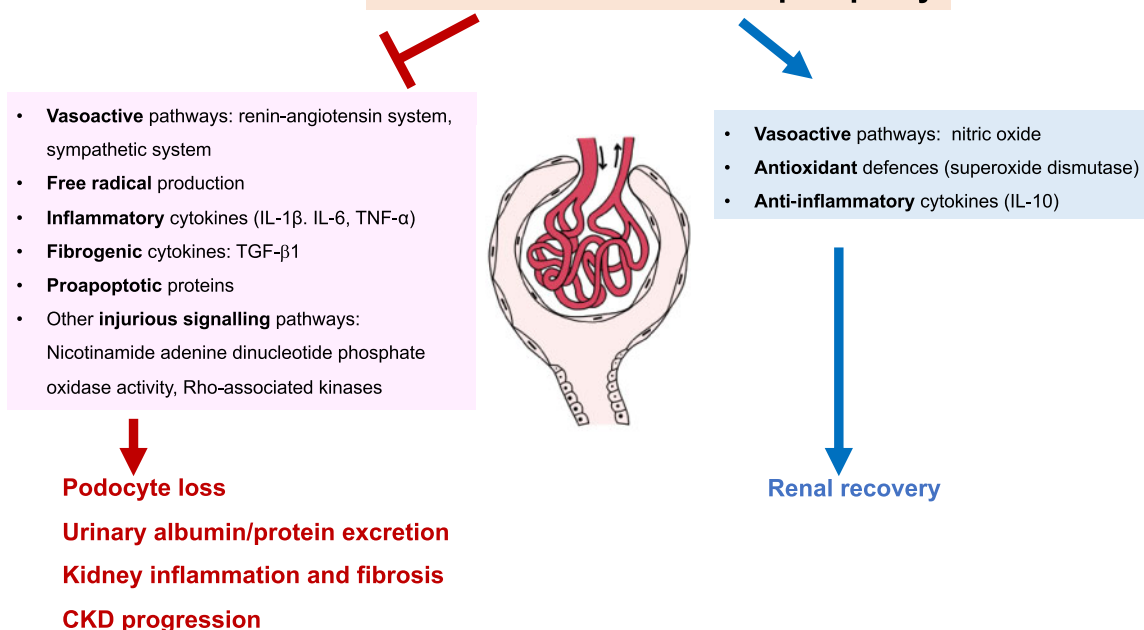


FIGURE 3: Postulated mechanisms of melatonin in the prevention and treatment of diabetic nephropathy. Melatonin may decrease podocyte loss, urinary albumin/protein excretion, kidney inflammation and fibrosis and may increase renal recovery.

associated kidney fibrosis [112]. Melatonin modulates macrophage recruitment and activation via multiple pathways, including NF- κ B activation [1]. Thus melatonin decreases M1 pro-inflammatory macrophage and increases M2 anti-inflammatory and reparative macrophages [113], blunting inflammatory cytokine secretion (IL-1 β , IL-6 and tumour necrosis factor α) and decreasing free radical production, while increasing the release of anti-inflammatory cytokine such as IL-10 from M2 macrophages [1].

Finally, it is important to correlate melatonin deficiency with obesity and hypertension since these are commonly discussed predisposing factors for DN. Obesity and hypertension frequently coexist [114] and are associated with oxidative stress and inflammation, especially at the vascular level. Specifically, kidney oxidative stress and inflammation contribute to hypertension [115].

As suggested above, melatonin has both anti-inflammatory and antioxidant effects due to cyclooxygenase synthase inhibition and multilevel inflammasome inhibition for cytokines, chemokines and adhesion molecules [116]. Melatonin decreases blood pressure via reduced NF- κ B activation and reduced renal inflammation in spontaneously hypertensive rats [117]. Qiao *et al.* [118] demonstrated that melatonin reduced hypertension and inflammatory cellular infiltration of the renal tubules.

Melatonin has many antioxidant effects. Those highlighted in the literature include reduction of oxidative stress, renal inflammation, proteinuria and progression of renal damage in rats with low renal mass [119]. Melatonin exerts renoprotective and anti-hypertensive effects by increasing nitric oxide bioavailability [120]. Melatonin deficiency is also related with obesity. Melatonin reduces body fat content, especially visceral fat, and improves metabolic condition via reduced free fatty acids, reduced hyperglycaemia and reduced insulin levels alongside improved high-density lipoprotein and adiponectin levels [25, 121–123].

It was shown that the amplitudes of the nocturnal pineal [124] and serum melatonin peaks decreased significantly in

obese animals. Daily melatonin supplementation significantly reduced body weight as well as plasma glucose, leptin, triglyceride and total cholesterol levels of the rat models of high-fat diet-induced obesity [125, 126]. The summary of evidence supports the hypothesis that melatonin deficiency plays a role in the development of kidney disease vis-à-vis obesity and hypertension.

CONCLUSION

Melatonin links sleep to metabolic and haemodynamic equilibrium. Melatonin activates the cardiovascular system and kidney receptors to protect from DN in preclinical models. Furthermore, melatonin levels are associated with human DN outcomes. Only human randomized controlled trials will confirm whether melatonin improves renal outcomes in diabetics and increases survival.

ACKNOWLEDGEMENTS

M.K. gratefully acknowledges use of the services and facilities of the Koç University Research Center for Translational Medicine (KUTTAM), funded by the Republic of Turkey Ministry of Development. The content is solely the responsibility of the authors and does not necessarily represent the official views of the Ministry of Development.

FUNDING

J.C.-N. is supported by FAPESP funding (2014/50457-0).

CONFLICT OF INTEREST STATEMENT

None declared.

REFERENCES

- Xia Y, Chen S, Zeng S et al. Melatonin in macrophage biology: current understanding and future perspectives. *J Pineal Res* 2019; 66: e12547
- Cipolla-Neto J, Amaral F. Melatonin as a hormone: new physiological and clinical insights. *Endocr Rev* 2018; 39: 990–1028
- Kato M, Natarajan R. Diabetic nephropathy—emerging epigenetic mechanisms. *Nat Rev Nephrol* 2014; 10: 517–530
- Zoungas S, Arima H, Gerstein HC et al. Effects of intensive glucose control on microvascular outcomes in patients with type 2 diabetes: a meta-analysis of individual participant data from randomised controlled trials. *Lancet Diabetes Endocrinol* 2017; 5: 431–437
- Kanwar YS, Sun L, Xie P et al. A glimpse of various pathogenetic mechanisms of diabetic nephropathy. *Annu Rev Pathol Mech Dis* 2011; 6: 395–423
- Pagtalunan ME, Miller PL, Jumping-Eagle S et al. Podocyte loss and progressive glomerular injury in type II diabetes. *J Clin Invest* 1997; 99: 342–348
- Lerner AB, Case JD, Mori W et al. Melatonin in peripheral nerve. *Nature* 1959; 183: 1821–1821
- Bubenik GA. Gastrointestinal melatonin: localization, function, and clinical relevance. *Dig Dis Sci* 2002; 47: 2336–2348
- Slominski A, Tobin DJ, Zmijewski MA et al. Melatonin in the skin: synthesis, metabolism and functions. *Trends Endocrinol Metab* 2008; 19: 17–24
- Canteras NS, Ribeiro-Barbosa ER, Goto M et al. The retinohypothalamic tract: comparison of axonal projection patterns from four major targets. *Brain Res Rev* 2011; 65: 150–183
- Drijfhout WJ, van der Linde AG, Kooi SE et al. Norepinephrine release in the rat pineal gland: the input from the biological clock measured by in vivo microdialysis. *J Neurochem* 2002; 66: 748–755
- Ma X, Idle JR, Krausz KW et al. Metabolism of melatonin by human cytochromes p450. *Drug Metab Dispos* 2005; 33: 489–494
- Tordjman S, Chokron S, Delorme R et al. Melatonin: pharmacology, functions and therapeutic benefits. *Curr Neuropsychopharmacol* 2017; 15: 434–443
- Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *J Pineal Res* 2013; 54: 245–257
- Rodriguez C, Mayo JC, Sainz RM et al. Regulation of antioxidant enzymes: a significant role for melatonin. *J Pineal Res* 2004; 36: 1–9
- Galano A, Medina ME, Tan DX et al. Melatonin and its metabolites as copper chelating agents and their role in inhibiting oxidative stress: a physicochemical analysis. *J Pineal Res* 2015; 58: 107–116
- Garcia JJ, Lopez-Pingarron L, Almeida-Souza P et al. Protective effects of melatonin in reducing oxidative stress and in preserving the fluidity of biological membranes: a review. *J Pineal Res* 2014; 56: 225–237
- Martin M, Macias M, Escames G et al. Melatonin-induced increased activity of the respiratory chain complexes I and IV can prevent mitochondrial damage induced by ruthenium red in vivo. *J Pineal Res* 2000; 28: 242–248
- Benitez-King G, Rios A, Martinez A et al. In vitro inhibition of Ca^{2+} /calmodulin-dependent kinase II activity by melatonin. *Biochim Biophys Acta* 1996; 1290: 191–196
- Karasek M, Winczyk K. Melatonin in humans. *J Physiol Pharmacol* 2006; 57(Suppl 5): 19–39
- Liu J, Clough SJ, Hutchinson AJ et al. MT1 and MT2 melatonin receptors: a therapeutic perspective. *Annu Rev Pharmacol Toxicol* 2016; 56: 361–383
- Pandi-Perumal SR, Srinivasan V, Maestroni GJ et al. Melatonin: nature's most versatile biological signal? *FEBS J* 2006; 273: 2813–2838
- Vimal RL, Pandey-Vimal MU, Vimal LS et al. Activation of suprachiasmatic nuclei and primary visual cortex depends upon time of day. *Eur J Neurosci* 2009; 29: 399–410
- Buonfiglio D, Parthimos R, Dantas R et al. Melatonin absence leads to long-term Leptin resistance and overweight in rats. *Front Endocrinol (Lausanne)* 2018; 9: 122
- Wolden-Hanson T, Mitton DR, McCants RL et al. Daily melatonin administration to middle-aged male rats suppresses body weight, intraabdominal adiposity, and plasma leptin and insulin independent of food intake and total body fat. *Endocrinology* 2000; 141: 487–497
- Zanuto R, Siqueira-Filho MA, Caperuto LC et al. Melatonin improves insulin sensitivity independently of weight loss in old obese rats. *J Pineal Res* 2013; 55: 156–165
- Amstrup AK, Sikjaer T, Pedersen SB et al. Reduced fat mass and increased lean mass in response to 1 year of melatonin treatment in postmenopausal women: a randomized placebo-controlled trial. *Clin Endocrinol* 2016; 84: 342–347
- Nishiyama K, Hirai K. The melatonin agonist ramelteon induces duration-dependent clock gene expression through cAMP signaling in pancreatic INS-1 β -cells. *PLoS One* 2014; 9: e102073
- Costes S, Boss M, Thomas AP, Matveyenko AV. Activation of melatonin signaling promotes beta-cell survival and function. *Mol Endocrinol* 2015; 29: 682–692
- Gil-Lozano M, Wu WK, Martchenko A, Brubaker PL. High-fat diet and palmitate alter the rhythmic secretion of glucagon-like peptide-1 by the rodent L-cell. *Endocrinology* 2016; 157: 586–599
- Kemp DM, Ubeda M, Habener JF. Identification and functional characterization of melatonin Mel 1a receptors in pancreatic beta cells: potential role in incretin-mediated cell function by sensitization of cAMP signaling. *Mol Cell Endocrinol* 2002; 191: 157–166
- Lindgren O, Mari A, Deacon CF et al. Differential islet and incretin hormone responses in morning versus afternoon after standardized meal in healthy men. *J Clin Endocrinol Metab* 2009; 94: 2887–2892
- Gil-Lozano M, Hunter PM, Behan LA et al. Short-term sleep deprivation with nocturnal light exposure alters time-dependent glucagon-like peptide-1 and insulin secretion in male volunteers. *Am J Physiol Endocrinol Metab* 2016; 310: E41–E50
- Rubio-Sastre P, Scheer FA, Gomez-Abellan P et al. Acute melatonin administration in humans impairs glucose tolerance in both the morning and evening. *Sleep* 2014; 37: 1715–1719
- McMullan CJ, Schernhammer ES, Rimm EB et al. Melatonin secretion and the incidence of type 2 diabetes. *JAMA* 2013; 309: 1388–1396
- Peliciari-Garcia RA, Marcal AC, Silva JA, Carmo-Buonfiglio D et al. Insulin temporal sensitivity and its signaling pathway in the rat pineal gland. *Life Sci* 2010; 87: 169–174
- Li HL, Kang YM, Yu L et al. Melatonin reduces blood pressure in rats with stress-induced hypertension via GABAA receptors. *Clin Exp Pharmacol Physiol* 2009; 36: 436–440
- Simko F, Reiter RJ, Pechanova O, Paulis L. Experimental models of melatonin-deficient hypertension. *Front Biosci* 2013; 18: 616–625

39. Viswanathan M, Laitinen JT, Saavedra JM. Vascular melatonin receptors. *Biol Signals* 1993; 2: 221–227
40. Baltatu OC, Amaral FG, Campos LA, Cipolla-Neto J. Melatonin, mitochondria and hypertension. *Cell Mol Life Sci* 2017; 74: 3955–3964
41. Wu YH, Zhou JN, Balesar R et al. Distribution of MT1 melatonin receptor immunoreactivity in the human hypothalamus and pituitary gland: colocalization of MT1 with vasopressin, oxytocin, and corticotropin-releasing hormone. *J Comp Neurol* 2006; 499: 897–910
42. Paulis L, Simko F. Blood pressure modulation and cardiovascular protection by melatonin: potential mechanisms behind. *Physiol Res* 2007; 56: 671–684
43. Campos LA, Cipolla-Neto J, Michelini LC. Melatonin modulates baroreflex control via area postrema. *Brain Behav* 2013; 3: 171–177
44. Patel KP, Li YF, Hirooka Y. Role of nitric oxide in central sympathetic outflow. *Exp Biol Med (Maywood)* 2001; 226: 814–824
45. Pechanova O, Paulis L, Simko F. Peripheral and central effects of melatonin on blood pressure regulation. *Int J Mol Sci* 2014; 15: 17920–17937
46. Simko F, Baka T, Paulis L, Reiter RJ. Elevated heart rate and nondipping heart rate as potential targets for melatonin: a review. *J Pineal Res* 2016; 61: 127–137
47. Jonas M, Garfinkel D, Zisapel N et al. Impaired nocturnal melatonin secretion in non-dipper hypertensive patients. *Blood Press* 2003; 12: 19–24
48. Cagnacci A, Cannoletta M, Renzi A et al. Prolonged melatonin administration decreases nocturnal blood pressure in women. *Am J Hypertens* 2005; 18: 1614–1618
49. Thomas MC, Brownlee M, Susztak K et al. Diabetic kidney disease. *Nat Rev Dis Primers* 2015; 1: 15018
50. Groop PH, Thomas MC, Moran JL et al. The presence and severity of chronic kidney disease predicts all-cause mortality in type 1 diabetes. *Diabetes* 2009; 58: 1651–1658
51. Afkarian M, Sachs MC, Kestenbaum B et al. Kidney disease and increased mortality risk in type 2 diabetes. *J Am Soc Nephrol* 2013; 24: 302–308
52. Mogensen CE, Christensen CK, Vittinghus E. The stages in diabetic renal disease. With emphasis on the stage of incipient diabetic nephropathy. *Diabetes* 1983; 32(Suppl 2): 64–78
53. Thomas MC, Macisaac RJ, Jerums G et al. Nonalbuminuric renal impairment in type 2 diabetic patients and in the general population (national evaluation of the frequency of renal impairment co-existing with NIDDM [NEFRON] 11). *Diabetes Care* 2009; 32: 1497–1502
54. Inoguchi T, Li P, Umeda F et al. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 2000; 49: 1939–1945
55. Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature* 2001; 414: 813–820
56. Cooper ME. Pathogenesis, prevention, and treatment of diabetic nephropathy. *Lancet* 1998; 352: 213–219
57. Derby L, Warram JH, Laffel LM et al. Elevated blood pressure predicts the development of persistent proteinuria in the presence of poor glycemic control, in patients with type I diabetes. *Diabetes Metab* 1989; 15: 320–326
58. Tanaka Y, Atsumi Y, Matsuo K et al. Role of glycemic control and blood pressure in the development and progression of nephropathy in elderly Japanese NIDDM patients. *Diabetes Care* 1998; 21: 116–120
59. Parving HH, Kasstrup H, Smidt UM et al. Impaired autoregulation of glomerular filtration rate in type 1 (insulin-dependent) diabetic patients with nephropathy. *Diabetologia* 1984; 27: 547–552
60. Christensen PK, Hansen HP, Parving HH. Impaired autoregulation of GFR in hypertensive non-insulin dependent diabetic patients. *Kidney Int* 1997; 52: 1369–1374
61. Blantz RC. Phenotypic characteristics of diabetic kidney involvement. *Kidney Int* 2014; 86: 7–9
62. UK Prospective Diabetes Study Group. Tight blood pressure control and risk of macrovascular and microvascular complications in type 2 diabetes: UKPDS 38. *BMJ* 1998; 317: 703–713
63. de Galan BE, Perkovic V, Ninomiya T et al. Lowering blood pressure reduces renal events in type 2 diabetes. *J Am Soc Nephrol* 2009; 20: 883–892
64. Lewis EJ, Hunsicker LG, Bain RP et al. The effect of angiotensin-converting-enzyme inhibition on diabetic nephropathy. *N Engl J Med* 1993; 329: 1456–1462
65. Parvanova AI, Trevisan R, Iliev IP et al. Insulin resistance and microalbuminuria: a cross-sectional, case-control study of 158 patients with type 2 diabetes and different degrees of urinary albumin excretion. *Diabetes* 2006; 55: 1456–1462
66. Thorn LM, Forsblom C, Fagerudd J et al. Metabolic syndrome in type 1 diabetes: association with diabetic nephropathy and glycemic control (the FinnDiane study). *Diabetes Care* 2005; 28: 2019–2024
67. Coward R, Fornoni A. Insulin signaling: implications for podocyte biology in diabetic kidney disease. *Curr Opin Nephrol Hypertens* 2015; 24: 104–110
68. Groop PH, Forsblom C, Thomas MC. Mechanisms of disease: pathway-selective insulin resistance and microvascular complications of diabetes. *Nat Rev Endocrinol* 2005; 1: 100–110
69. Chow FY, Nikolic-Paterson DJ, Ozols E et al. Monocyte chemoattractant protein-1 promotes the development of diabetic renal injury in streptozotocin-treated mice. *Kidney Int* 2006; 69: 73–80
70. Lim AK, Tesch GH. Inflammation in diabetic nephropathy. *Mediators Inflamm* 2012; 2012: 1–12
71. Chow FY, Nikolic-Paterson DJ, Ozols E et al. Intercellular adhesion molecule-1 deficiency is protective against nephropathy in type 2 diabetic db/db mice. *J Am Soc Nephrol* 2005; 16: 1711–1722
72. Kanamori H, Matsubara T, Mima A et al. Inhibition of MCP-1/CCR2 pathway ameliorates the development of diabetic nephropathy. *Biochem Biophys Res Commun* 2007; 360: 772–777
73. Tutuncu NB, Batur MK, Yildirim A et al. Melatonin levels decrease in type 2 diabetic patients with cardiac autonomic neuropathy. *J Pineal Res* 2005; 39: 43–49
74. Hikichi T, Tateda N, Miura T. Alteration of melatonin secretion in patients with type 2 diabetes and proliferative diabetic retinopathy. *Clin Ophthalmol* 2011; 5: 655–660
75. O'Brien IA, Lewin IG, O'Hare JP et al. Abnormal circadian rhythm of melatonin in diabetic autonomic neuropathy. *Clin Endocrinol (Oxf)* 1986; 24: 359–364
76. Patel SR, Hu FB. Short sleep duration and weight gain: a systematic review. *Obesity (Silver Spring)* 2008; 16: 643–653
77. Reiter RJ, Tan DX, Korkmaz A et al. Obesity and metabolic syndrome: association with chronodisruption, sleep deprivation, and melatonin suppression. *Ann Med* 2012; 44: 564–577

78. Arora T, Chen MZ, Cooper AR et al. The impact of sleep debt on excess adiposity and insulin sensitivity in patients with early type 2 diabetes mellitus. *J Clin Sleep Med* 2016; 12: 673–680
79. Xie Z, Chen F, Li WA et al. A review of sleep disorders and melatonin. *Neurol Res* 2017; 39: 559–565
80. Afsar B. The relationship between self-reported nocturnal sleep duration, daytime sleepiness and 24-h urinary albumin and protein excretion in patients with newly diagnosed type 2 diabetes. *Prim Care Diabetes* 2013; 7: 39–44
81. Amaral FG, Turati AO, Barone M et al. Melatonin synthesis impairment as a new deleterious outcome of diabetes-derived hyperglycemia. *J Pineal Res* 2014; 57: 67–79
82. Peschke E, Frese T, Chankiewicz E et al. Diabetic Goto Kakizaki rats as well as type 2 diabetic patients show a decreased diurnal serum melatonin level and an increased pancreatic melatonin-receptor status. *J Pineal Res* 2006; 40: 135–143
83. Kor Y, Geyikli I, Keskin M et al. Preliminary study: evaluation of melatonin secretion in children and adolescents with type 1 diabetes mellitus. *Indian J Endocr Metab* 2014; 18: 565–568
84. Robeva R, Kirilov G, Tomova A et al. Melatonin-insulin interactions in patients with metabolic syndrome. *J Pineal Res* 2008; 44: 52–56
85. Lai AY, McLaurin J. Rho-associated protein kinases as therapeutic targets for both vascular and parenchymal pathologies in Alzheimer's disease. *J Neurochem* 2018; 144: 659–668
86. Korol A, Taiyab A, West-Mays JA. RhoA/ROCK signaling regulates TGF β -induced epithelial-mesenchymal transition of lens epithelial cells through MRTF-A. *Mol Med* 2016; 22: 713–723
87. Kolavennu V, Zeng L, Peng H et al. Targeting of RhoA/ROCK signaling ameliorates progression of diabetic nephropathy independent of glucose control. *Diabetes* 2008; 57: 714–723
88. Komers R, Oyama TT, Beard DR et al. Rho kinase inhibition protects kidneys from diabetic nephropathy without reducing blood pressure. *Kidney Int* 2011; 79: 432–442
89. Liu F, Zhang S, Xu R et al. Melatonin attenuates endothelial-to-mesenchymal transition of glomerular endothelial cells via regulating miR-497/ROCK in diabetic nephropathy. *Kidney Blood Press Res* 2018; 43: 1425–1436
90. Rashed LA, Elattar S, Eltablawy N et al. Mesenchymal stem cells pretreated with melatonin ameliorate kidney functions in a rat model of diabetic nephropathy. *Biochem Cell Biol* 2018; 96: 564–571
91. Yacoub R, Campbell KN. Inhibition of RAS in diabetic nephropathy. *Int J Nephrol Renovasc Dis* 2015; 8: 29–40
92. Miyata K, Ohashi N, Suzuki Y et al. Sequential activation of the reactive oxygen species/angiotensinogen/renin-angiotensin system axis in renal injury of type 2 diabetic rats. *Clin Exp Pharmacol Physiol* 2008; 35: 922–927
93. Kamiyama M, Urushihara M, Morikawa T et al. Oxidative stress/angiotensinogen/renin-angiotensin system axis in patients with diabetic nephropathy. *Int J Mol Sci* 2013; 14: 23045–23062
94. Ishigaki S, Ohashi N, Isobe S et al. Impaired endogenous nighttime melatonin secretion relates to intrarenal renin-angiotensin system activation and renal damage in patients with chronic kidney disease. *Clin Exp Nephrol* 2016; 20: 878–884
95. Ishigaki S, Ohashi N, Matsuyama T et al. Melatonin ameliorates intrarenal renin-angiotensin system in a 5/6 nephrectomy rat model. *Clin Exp Nephrol* 2018; 22: 539–549
96. Ji ZZ, Xu YC. Melatonin protects podocytes from angiotensin II-induced injury in an in vitro diabetic nephropathy model. *Mol Med Rep* 2016; 14: 920–926
97. Ha H, Yu MR, Kim KH. Melatonin and taurine reduce early glomerulopathy in diabetic rats. *Free Radic Biol Med* 1999; 26: 944–950
98. Anwar MM, Meki AR. Oxidative stress in streptozotocin-induced diabetic rats: effects of garlic oil and melatonin. *Comp Biochem Physiol A Mol Integr Physiol* 2003; 135: 539–547
99. Motawi TK, Ahmed SA, Hamed MA et al. Combination of melatonin and certain drugs for treatment of diabetic nephropathy in streptozotocin-induced diabetes in rats. *Diabetol Int* 2016; 7: 413–424
100. Elbe H, Vardi N, Esrefoglu M et al. Amelioration of streptozotocin-induced diabetic nephropathy by melatonin, quercetin, and resveratrol in rats. *Hum Exp Toxicol* 2015; 34: 100–113
101. Onk D, Onk OA, Turkmen K et al. Melatonin attenuates contrast-induced nephropathy in diabetic rats: the role of interleukin-33 and oxidative stress. *Mediators Inflamm* 2016; 2016: 1–10
102. Aygun H, Gul SS. Protective effect of melatonin and agomelatine on adriamycin-induced nephrotoxicity in rat model: a renal scintigraphy and biochemical study. *Bratisl Lek Listy* 2019; 120: 113–118
103. Gorin Y, Block K. Nox as a target for diabetic complications. *Clin Sci (Lond)* 2013; 125: 361–382
104. Altenhofer S, Kleikers PW, Radermacher KA et al. The NOX toolbox: validating the role of NADPH oxidases in physiology and disease. *Cell Mol Life Sci* 2012; 69: 2327–2343
105. Winiarska K, Dzik JM, Labudda M et al. Melatonin nephroprotective action in Zucker diabetic fatty rats involves its inhibitory effect on NADPH oxidase. *J Pineal Res* 2016; 60: 109–117
106. Yang H, Xie T, Li D et al. Tim-3 aggravates podocyte injury in diabetic nephropathy by promoting macrophage activation via the NF- κ B/TNF- α pathway. *Mol Metab* 2019; 23: 24–36
107. Sassy-Prigent C, Heudes D, Mandet C et al. Early glomerular macrophage recruitment in streptozotocin-induced diabetic rats. *Diabetes* 2000; 49: 466–475
108. Chow F, Ozols E, Nikolic-Paterson DJ et al. Macrophages in mouse type 2 diabetic nephropathy: correlation with diabetic state and progressive renal injury. *Kidney Int* 2004; 65: 116–128
109. You H, Gao T, Cooper TK et al. Macrophages directly mediate diabetic renal injury. *Am J Physiol Renal Physiol* 2013; 305: F1719–F1727
110. de Zeeuw D, Bekker P, Henkel E et al. The effect of CCR2 inhibitor CCX140-B on residual albuminuria in patients with type 2 diabetes and nephropathy: a randomised trial. *Lancet Diabetes Endocrinol* 2015; 3: 687–696
111. Caamano J, Hunter CA. NF- κ B family of transcription factors: central regulators of innate and adaptive immune functions. *Clin Microbiol Rev* 2002; 15: 414–429
112. Pawluczyk IZ, Harris KP. Macrophages promote pro-sclerotic responses in cultured rat mesangial cells: a mechanism for the initiation of glomerulosclerosis. *J Am Soc Nephrol* 1997; 8: 1525–1536
113. Yi WJ, Kim TS. Melatonin protects mice against stress-induced inflammation through enhancement of M2 macrophage polarization. *Int Immunopharmacol* 2017; 48: 146–158

114. DeMarco VG, Aroor AR, Sowers JR. The pathophysiology of hypertension in patients with obesity. *Nat Rev Endocrinol* 2014; 10: 364–376
115. Prado NJ, Ferder L, Manucha W, Diez ER. Anti-inflammatory effects of melatonin in obesity and hypertension. *Curr Hypertens Rep* 2018; 20: 45
116. Szewczyk-Golec K, Wozniak A, Reiter RJ. Inter-relationships of the chronobiotic, melatonin, with leptin and adiponectin: implications for obesity. *J Pineal Res* 2015; 59: 277–291
117. Nava M, Quiroz Y, Vaziri N et al. Melatonin reduces renal interstitial inflammation and improves hypertension in spontaneously hypertensive rats. *Am J Physiol Renal Physiol* 2003; 284: F447–F454
118. Qiao YF, Guo WJ, Li L et al. Melatonin attenuates hypertension-induced renal injury partially through inhibiting oxidative stress in rats. *Mol Med Rep* 2016; 13: 21–26
119. Quiroz Y, Ferrebuz A, Romero F et al. Melatonin ameliorates oxidative stress, inflammation, proteinuria, and progression of renal damage in rats with renal mass reduction. *Am J Physiol Renal Physiol* 2008; 294: F336–F344
120. Cheng MC, Wu TH, Huang LT et al. Renoprotective effects of melatonin in young spontaneously hypertensive rats with L-NAME. *Pediatr Neonatol* 2014; 55: 189–195
121. Nduhirabandi F, Du Toit EF, Lochner A. Melatonin and the metabolic syndrome: a tool for effective therapy in obesity-associated abnormalities? *Acta Physiol* 2012; 205: 209–223
122. Rasmussen DD, Boldt BM, Wilkinson CW et al. Daily melatonin administration at middle age suppresses male rat visceral fat, plasma leptin, and plasma insulin to youthful levels. *Endocrinology* 1999; 140: 1009–1012
123. Nishida S. Metabolic effects of melatonin on oxidative stress and diabetes mellitus. *Endocrine* 2005; 27: 131–136
124. Cano P, Jimenez-Ortega V, Larrad A et al. Effect of a high-fat diet on 24-h pattern of circulating levels of prolactin, luteinizing hormone, testosterone, corticosterone, thyroid-stimulating hormone and glucose, and pineal melatonin content, in rats. *Endocrine* 2008; 33: 118–125
125. Prunet-Marcassus B, Desbazeille M, Bros A et al. Melatonin reduces body weight gain in Sprague Dawley rats with diet-induced obesity. *Endocrinology* 2003; 144: 5347–5352
126. Rios-Lugo MJ, Cano P, Jimenez-Ortega V et al. Melatonin effect on plasma adiponectin, leptin, insulin, glucose, triglycerides and cholesterol in normal and high fat-fed rats. *J Pineal Res* 2010; 49: 342–348